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Nextera® XT DNA Library Preparation Kit

The fastest and easiest library prep workflow for small genomes, PCR amplicons, and plasmids.

Highlights

- **Rapid Library Preparation**
  Complete library prep in as little as 90 minutes with only 15 minutes of hands-on time

- **Fastest Time to Results**
  Go from DNA to data in 8 hours with the MiSeq® System

- **Optimized for Small Genomes, PCR Amplicons, and Plasmids**
  One library prep kit for many applications

- **Innovative Sample Normalization**
  Eliminates the need for library quantification before sample pooling and sequencing

Introduction

The Nextera XT DNA Library Preparation Kit enables researchers to prepare sequencing-ready libraries for small genomes (bacteria, archaea, and viruses), PCR amplicons, and plasmids in 90 minutes, with only 15 minutes of hands-on time. The combination of the MiSeq System and Nextera XT DNA Library Preparation Kits enable you to go from DNA to data in 8 hours (Figure 1). The low amount (1 ng) of input DNA makes this method amenable to precious samples available in limited quantity. Compatible with all Illumina sequencers, Nextera library preparation can shorten the overall sequencing workflow time for a wide variety of established applications[1-9] and can be automated easily for greater throughput.

Fastest and Easiest Library Prep Workflow

Using a single "tagmentation" enzymatic reaction, sample DNA is simultaneously fragmented and tagged with adapters. An optimized, limited-cycle PCR protocol amplifies tagged DNA and adds sequencing indexes (Figure 1). From start to finish, the complete Nextera XT protocol is over 80% faster than other available library preparation methods, and requires the least amount of hands-on time.

Innovative Sample Normalization

Library preparation kits for next-generation sequencing result in libraries of varying concentration. To pool samples equally and achieve target cluster densities, time-intensive quantitation methods are often used, followed by dilution and pooling of barcoded samples. The Nextera XT DNA Library Preparation Kit eliminates the need for library quantification before sample pooling and sequencing by employing a simple bead-based sample normalization step (Figure 2). Prepared libraries are produced at equivalent concentrations enabling pooling by volume—simply pool 5 µl of each library to be sequenced.

Flexible Multiplexing

The Nextera XT Library Preparation Kit features an innovative indexing solution for processing and uniquely barcoding up to 384 samples in a single experiment. Following the addition of two indexes to each DNA fragment, up to 384 uniquely indexed samples can be pooled and sequenced together. After sequencing, the unique combination of the two indexes is used to demultiplex the data and assign reads to the proper sample. Using this dual-barcode approach, Nextera XT Index Kits only require 40 unique index oligos to process up to 384 samples
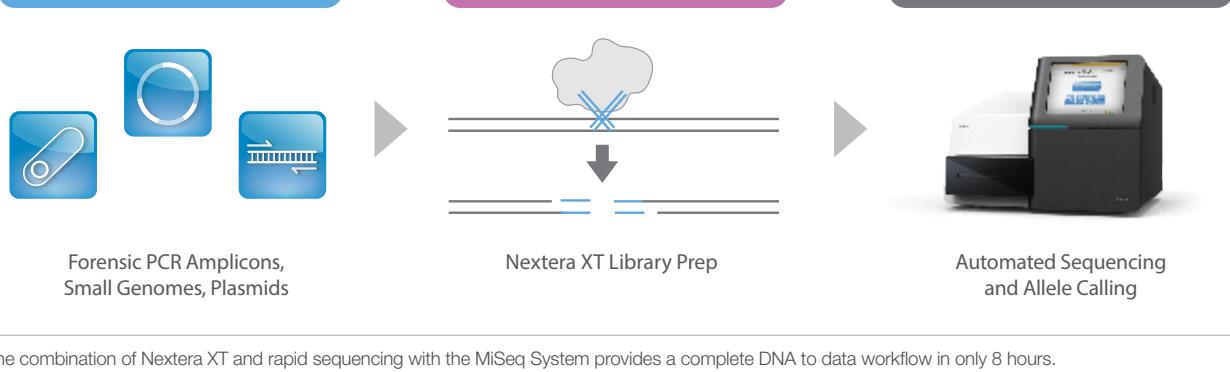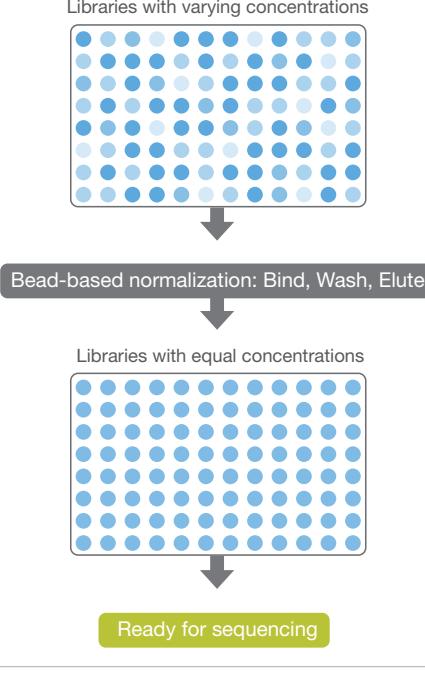
Figure 1: Nextera XT Library Preparation Workflow

Prepare Input DNA (1 ng) → Nextera Tagmentation → Sequencing and Analysis

Forensic PCR Amplicons, Small Genomes, Plasmids — Nextera XT Library Prep — Automated Sequencing and Allele Calling

The combination of Nextera XT and rapid sequencing with the MiSeq System provides a complete DNA to data workflow in only 8 hours.

Figure 2: Innovative Sample Normalization

Libraries with varying concentrations

Bead-based normalization: Bind, Wash, Elute

Libraries with equal concentrations

Ready for sequencing

The Nextera XT Library Preparation kit eliminates the need for library quantification before sample pooling and sequencing. Libraries of equivalent concentrations are created by employing bead-based sample normalization, as simple as pipetting 5 µl of each library to be sequenced.

for a scalable approach. Multisample studies can be conveniently managed using the Illumina Experiment Manager, a freely available software tool that provides easy reaction setup for plate-based processing.

Simple User Interface for Analysis

MiSeq Reporter provides automated on-instrument analysis for various applications including small genome *de novo* or resequencing, PCR amplicon, and plasmid sequencing. Sequencing results and analysis are easy to view and interpret. For example, using the PCR Amplicon workflow in the MiSeq Reporter software, sequence data are automatically categorized into intuitive tabs: Samples, Regions, and Variants (Figure 3). Within each of these tabs, the variant score, quality (Q) score, and sequencing coverage levels can be determined down to single bases, allowing easy analysis of variants of interest.

High Coverage, Accurate Calls

To illustrate the power of amplicon sequencing with Nextera XT and the MiSeq System, nine PCR amplicons of varying sizes were prepared from two different samples of human DNA. Amplicons from each sample were pooled and 1 ng of DNA from each pool was prepared using the Nextera XT kit. Libraries from the two sample pools were combined, sequenced with paired-end 2 × 150 reads on the MiSeq System, and analyzed with MiSeq Reporter using the PCR Amplicon workflow. The approximate mean sequencing coverage values per amplicon and number of variants called (variant score > 99) identified within the amplicons in one of the two samples are shown in Table 1. The output of the MiSeq System supported sequencing of these amplicons to a depth of > 12,000×, enabling

Figure 3: PCR Amplicon Workflow in MiSeq Reporter

MiSeq Reporter provides automated on-instrument analysis for various applications, including PCR amplicon shown here. Samples, regions, and variants are easily accessible, and variant scores, quality (Q) scores, and coverage plots are shown at single nucleotide resolution.
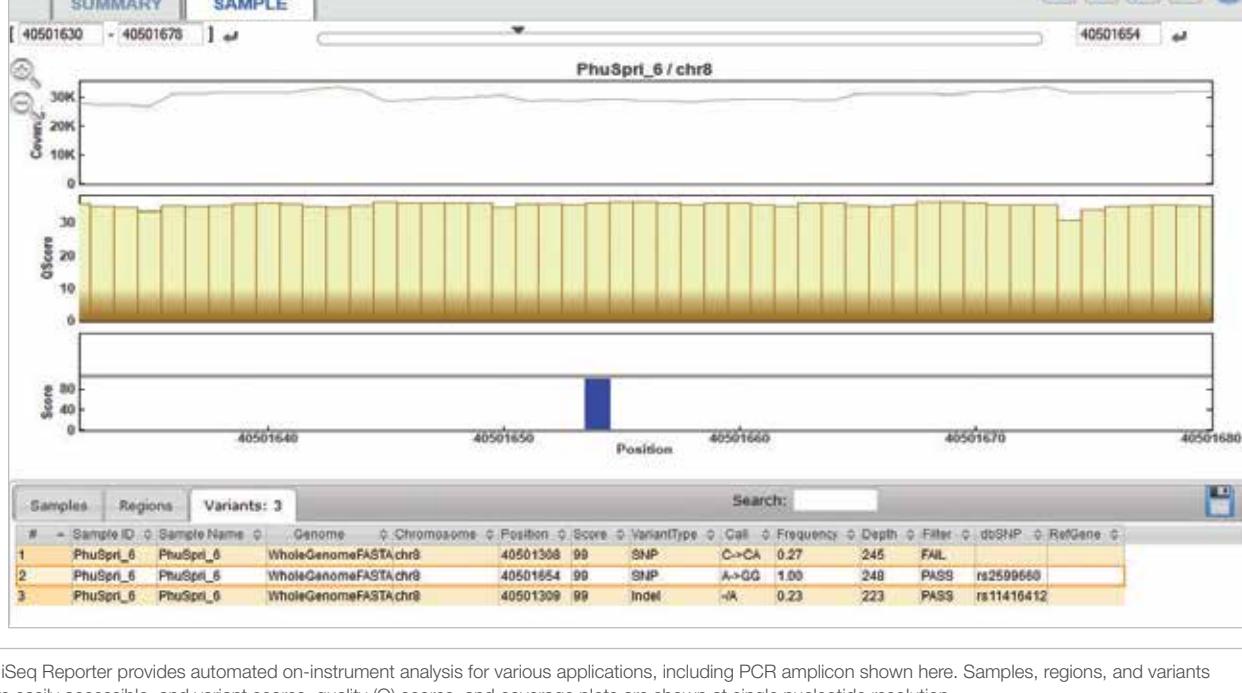
Table 1: Amplicon Coverage and Variants Called

Amplicon Length (bp)	Mean Coverage (thousands of reads)	Variants Called (SNVs/Indels)
953	15.1	4 SNVs
1083	27.4	4 SNVs
1099	22.1	1 SNV
1800	22.4	7 SNVs
1809	17.8	1 SNV
2166	17.6	7 SNVs
3064	12.5	4 SNVs
3064	13.3	1 SNV
3072	14.8 K	1 SNV + 1 indel

confident variant calling. Of the 31 total variants called in this example, 94% are confirmed within the dbSNP database. These results show that coverage is high and even across a range of amplicon sizes, and that variant calls are accurate.

Even Coverage Across Large Amplicons

Large amplicons (> 1 kb) produced by long-range PCR can be easily prepared with the Nextera XT kit and sequenced on any Illumina sequencer. In Figure 4, coverage along amplicon length and position of called variants is shown for a single 5.1 kb amplicon in a highly variable non-coding region of the human genome. The 5.1 kb amplicon was part of a pool of 24 amplicons from human DNA ranging in size from ~300 bp up to 10 kb. Amplicon pools were generated from five different samples, and Nextera XT libraries were made using 1 ng of DNA from each pool. Libraries were combined and single-read sequencing was performed using 1 × 150 bp cycles on MiSeq and analyzed using MiSeq Reporter with the PCR Amplicon workflow.

De Novo Assembly of Small Genomes

To show the utility of Nextera XT for preparing microbial genomes, 1 ng of genomic DNA from *Escherichia coli* reference strain MG1655 was prepared using the Nextera XT kit and sequenced using paired-end 2 × 150 bp reads on the MiSeq System. The data were analyzed using the Assembly workflow on the MiSeq Reporter. Total post-run analysis time for this sample was 28 minutes. Assembly metrics are shown in Table 2. A high-quality assembly was produced, with excellent N50 scores and coverage. This data set is available for analysis in BaseSpace®, the Illumina cloud computing environment[10].

Figure 4: Coverage of Large Amplicons
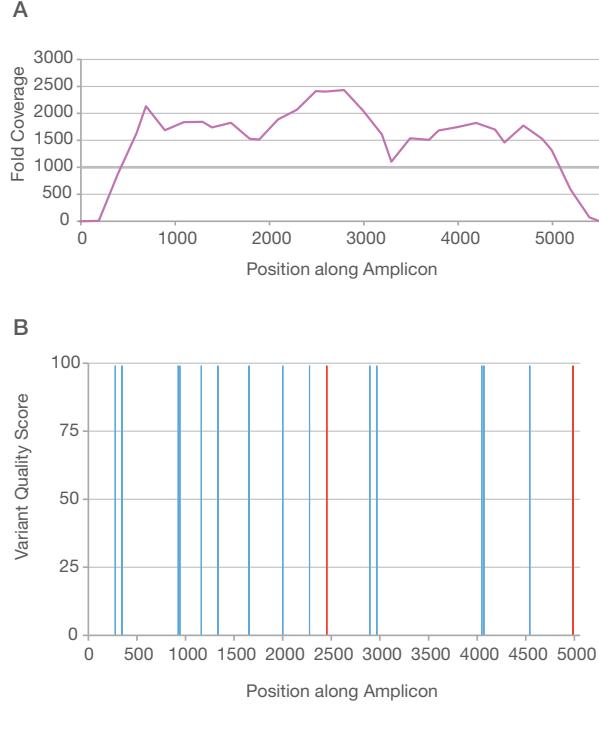
Panel A: High sequencing coverage (>1,000×) across a 5.1 kb amplicon
Panel B: Within the same amplicon, the position of 16 variants passing filter (14 SNVs in blue + 2 indels in red) is shown, plotted against variant score (a Phred-scaled measure of variant calling accuracy, maximum = 99). Of the 16 variants, 13 are present in dbSNP.

Table 2: *De Novo* Assembly of *E. coli*

Parameter	Value
Percent of genome covered	98%
Number of contigs	314
Maximum contig length	221,108
Base count	4,548,900
N50	111,546
Average coverage per base	184.9

Summary

Nextera XT DNA Library Preparation Kits are ideal for experiments where speed and ease are of paramount importance. Providing the fastest and easiest library preparation workflow, the Nextera XT DNA Library Preparation Kit enables rapid sequencing of small genomes, PCR amplicons, and plasmids. Combined with the MiSeq and NextSeq™ Systems, Nextera XT DNA Library Preparation Kits enable you to go from DNA to data—all in a single day.

References

1. Loman N, Misra RJ, Dallman TJ, Constantinidou C, Gharbia SE, et al. (2012) Performance comparison of benchtop high-throughput sequencing platforms. Nat Biotechnol 22 Apr.

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10. basespace.illumina.com

Nextera XT DNA Library Prep Kit Specifications

Specification	Value
Sample DNA input type	Genomic DNA, PCR amplicons, plasmids
Input DNA	1 ng
Typical median insert size	< 300 bp
Available indexes	Up to 384
Compatible sequencers	MiSeq, NextSeq, and HiSeq® Systems
Read lengths supported	Supports all read lengths on any Illumina sequencing system

Ordering Information

Product	Catalog No.
Nextera XT DNA Library Preparation Kit (24 samples)	FC-131-1024
Nextera XT DNA Library Preparation Kit (96 samples)	FC-131-1096
Nextera XT Index Kit (24 indexes, 96 samples)	FC-131-1001
Nextera XT Index Kit (96 indexes, 384 samples)	FC-131-1002
TruSeq® Dual Index Sequencing Primer Kit, Single Read (single-use kit)*	FC-121-1003
TruSeq Dual Index Sequencing Primer Kit, Paired-End Read (single-use kit)*	PE-121-1003
Nextera XT Index Kit v2, Set A (96 indexes, 384 samples)	FC-131-2001
Nextera XT Index Kit v2, Set B (96 indexes, 384 samples)	FC-131-2002
Nextera XT Index Kit v2, Set C (96 indexes, 384 samples)	FC-131-2003
Nextera XT Index Kit v2, Set D (96 indexes, 384 samples)	FC-131-2004

*Sequencing primer kits are required for all sequencers except the MiSeq System.

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